Metabolism of Synthetic Steroids by the Human Placenta

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Abstract

Pregnant women with asthma are frequently exposed to synthetic glucocorticoids and glucocorticoids are known to reduce fetal growth. The fetus is normally protected from the harmful effects of maternally derived glucocorticoids by the placental enzyme 11\textsubscript{b}-hydroxysteroid dehydrogenase type 2 (11\textsubscript{b}-HSD2). Whether 11\textsubscript{b}-HSD2 inactivates the synthetic glucocorticoids used for asthma treatment during pregnancy (budesonide, beclomethasone dipropionate and fluticasone propionate) remains unknown.

To investigate the relationship between steroid use during pregnancy and fetal growth and development, pregnant women with ($n = 119$) and without asthma ($n = 84$) were followed throughout pregnancy. Data on asthma medication use, neonatal size at birth, placental weight and cord blood cortisol and estriol were collected. Placental tissue samples were collected from non-asthmatic women ($n = 8$) for metabolism studies.

Placental 11\textsubscript{b}-HSD2 metabolised beclomethasone, prednisolone, dexamethasone and betamethasone, but not budesonide or fluticasone. No association between the use of inhaled steroids for asthma treatment during pregnancy and alterations in neonatal size, placental weight, gestational age at delivery, or umbilical vein estriol concentrations was demonstrated compared to non-asthmatic women. In conclusion, the use of inhaled steroids for asthma treatment does not affect fetal growth, despite differences in placental metabolism by 11\textsubscript{b}-HSD2.

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1. Introduction

Pregnant women with asthma are at increased risk of poor pregnancy outcomes [1], including preterm delivery [2], pre-eclampsia [3] and low birth weight [4] independent of prematurity [5]. The mechanisms contributing to these outcomes are unknown, but may include poorly controlled asthma [5] or its treatment with inhaled or oral corticosteroids [6]. While there is little evidence in the literature to suggest that inhaled corticosteroids (ICS) are harmful to the fetus [7,8], non-adherence to ICS medication by pregnant women with asthma for fear of its effect on the fetus remains a major clinical problem [9,10].

Glucocorticoids are essential for the development and maturation of fetal organs before birth and late pregnancy is characterised by a rise in cortisol levels, which parallels the increased maturity of fetal organs [11]. However, glucocorticoids have potentially adverse effects on the fetus and have been linked to altered fetal programming. Women using prednisone during pregnancy have been reported to have an
increase in still-birth, fetal distress, placental insufficiency [12] and low birth weight neonates [13]. Antenatal dexamethasone treatment has been associated with a reduction in birth weight, by as much as 161 g in infants delivered between 30 and 32 weeks [14]. While recent evidence from randomised controlled trials suggests that there is no additional decrease in fetal growth when repeated courses of antenatal steroids are used compared to single doses [15], French et al. found that repeated courses of betamethasone were associated with a 9% reduction in birth weight and a 4% reduction in head circumference in preterm infants born prior to 33 weeks of gestation [16].

Two isoforms of 11β-hydroxysteroid dehydrogenase (11β-HSD) have been cloned and characterised in humans which interconvert glucocorticoids with their 11-keto metabolites (Fig. 1). The type 1 enzyme (11β-HSD1) is NADP(H) dependent, acting primarily as an oxoreductase, converting cortisol to cortisone. The main functional isozyme in the placental syncytiotrophoblast is 11β-HSD2, a high affinity, NAD dependent, uni-directional enzyme, catalysing only the dehydrogenase reaction, converting cortisol to cortisone [17]. In the placenta 11β-HSD2 protects the fetus from the potentially harmful effects of endogenous maternal glucocorticoids [18], by acting as a barrier to protect the fetus from the much higher levels of cortisol found in the mother [19].

Several studies have described a relationship between reduced activity of 11β-HSD2 and reduced birth weight or intrauterine growth restriction [20,21]. We have previously found that asthmatic women who do not use inhaled steroids for treatment have smaller female neonates at term [22]. This late gestation reduction in fetal growth was associated with a significant decrease in 11β-HSD2 activity [22,23].

It is thought that synthetic steroid derivatives, such as dexamethasone and betamethasone are not extensively metabolised by 11β-HSD2. In bronchial epithelial cells, Feinstein and Schleimer found that the addition of the 11β-HSD inhibitor, glycyrrhetinic acid, had no effect on the anti-inflammatory activity of beclomethasone dipropionate (beclomethasone), fluticasone propionate (fluticasone) or budesonide, suggesting that they were not metabolised by 11β-HSD2 [24]. However, in another study using BEAS-2B cells derived from the bronchial epithelium, the addition of the 11β-HSD inhibitor, carbopenoxalone, resulted in an increased potency of dexamethasone by approximately 10-fold [25]. Orsida et al. have suggested that the presence of 11β-HSD2 activity in the lung may serve to re-activate synthetic steroids used as anti-inflammatory agents in diseases such as asthma [26]. Information about the metabolism of synthetic steroid derivatives by 11β-HSD2 is therefore conflicting.

No studies have been published regarding the placental metabolism of synthetic steroids used for asthma treatment during pregnancy such as beclomethasone, budesonide and fluticasone. If these steroids reach the placenta but are not metabolised into inactive forms, they may have adverse effects on fetal development and may contribute to reduced fetal growth in asthmatic women using inhaled or oral corticosteroid medication. We have studied metabolism of the synthetic steroids in placentae collected from non-asthmatic women, since changes in enzyme activity were previously observed in some women with asthma [22,23]. The effect of inhaled steroid use in vivo has been investigated in women with asthma, compared to women without asthma, by examining fetal growth and markers of fetal hypothalamic–pituitary–adrenal (HPA) axis function, including cortisol and estriol concentrations in cord blood. Estriol is a derivative of dehydroepiandrosterone sulfate (DHEA-S), derived from the fetal adrenal and produced by the placenta [27]. Less than 10% of circulating estriol is derived from the mother, estriol had been used as an indicator of fetal adrenal function and fetal well-being [27,28]. In this paper, we hypothesise that beclomethasone, budesonide, fluticasone, dexamethasone, betamethasone and prednisolone are converted into their 11-keto metabolites by the placental enzyme 11β-HSD2 and that the use of inhaled steroids by pregnant women with asthma has no effect on fetal growth and development.

2. Materials and methods

The studies were approved by the Hunter Area Health Service and University of Newcastle Human Research Ethics Committees. Pregnant women with and without asthma were recruited at the John Hunter Hospital antenatal clinics as part of a prospective cohort study, according to a previously described protocol [22,23,29]. We have previously reported some maternal characteristics, maternal corticosteroid use and some placental and fetal outcomes for a number of these subjects [22,23,29,30]. Neonatal size at birth (weight, length and head circumference) was recorded in 84 pregnant women without asthma and 119 pregnant women with asthma who used beclomethasone alone (\( n = 29 \)), budesonide alone (\( n = 42 \)), fluticasone alone (\( n = 31 \)) or inhaled corticosteroid (ICS) plus periodic oral steroid (prednisone, \( n = 17 \)) to treat their asthma during pregnancy. The dose of ICS used by the mothers in each trimester was recorded and summarised as the mean daily dose of beclomethasone dipropionate or equivalent, where 1 µg beclomethasone was considered equal

![Fig. 1. The reaction catalysed by placental 11β-HSD2.](image-url)
to 1 μg budesonide and 0.5 μg fluticasone [31]. The dose and length of use of oral steroids were not recorded. No women were using long acting β₂-agonists. Gestational age was calculated from the date of the last menstrual period and the 18-week ultrasound. Preterm delivery was defined as delivery at <37 completed weeks of gestation. In a subset of samples, placental weight was recorded and fetal HPA function was examined using measurements of umbilical vein cortisol and estriol as previously described [22].

2.1. Metabolism studies – placenta collection

Pregnant women who did not have diabetes, hypertension or asthma and were non-smokers were recruited by midwives from John Hunter Hospital and provided written informed consent for participation. All placentae (n = 8) were collected from term, uncomplicated vaginal deliveries. The placenta was collected within 45 min of delivery and pieces were snap frozen in liquid nitrogen and stored at −80 °C until further use.

2.2. Protein extraction

Approximately 1–2 g of snap frozen placenta was crushed with a mortar and pestle under liquid nitrogen. Crushed placental tissue was homogenised in 10 volumes of 0.1 M sodium phosphate buffer (pH 7.4) containing protease inhibitors (2 mM ethylenediamine tetra-acetic acid, protease inhibitor cocktail tablets 2/1, 5000 IU/l trasylo, 100 μM dithiothreitol, 1 μM pepstatin A, 1 mM benzamidine, 65 000 U/l bacitracin) and 0.25 M sucrose, using a polytron homogeniser (Kinematica AG, Switzerland). The homogenate was centrifuged at 1000 g for 15 min to remove cellular debris and the supernatant centrifuged at 100 000 g for 60 min to obtain the microsomal fraction. The pellet was re-homogenised in 2 ml sodium phosphate buffer containing protease inhibitors and the protein concentration determined by Bradford Assay [32] against a standard curve of bovine serum albumin.

2.3. Placental 11β-HSD2 activity assay

Placental 11β-HSD2 activity was measured using a modification of the method previously used in our laboratory [22,23,33]. Initially, protein concentration (100, 150 and 200 μg/ml), time (15 min, 1, 2, 4, 6 and 24 h) and steroid substrate concentration (5 and 50 μM) were optimised. In the final method used, 50 μM steroid substrate (beclomethasone, budesonide, fluticasone, prednisolone, dexamethasone or betamethasone) was chosen due to the need to visualise the steroid on the thin layer chromatography (TLC) plate. We have previously demonstrated that placental 11β-HSD2 is saturated by 5 μM cortisol [22,23]. Steroids were obtained from Sigma (St Louis, MO, USA), except for fluticasone which was reconstituted from the powder of 45 blisters of a Flutotide™ Accuhaler™ (each blister contained 500 μg fluticasone) obtained from the hospital pharmacy. The powder, 2.7 ml water was added and thoroughly mixed, in an attempt to remove the lactose. The mixture was centrifuged at 1000 g for 10 min and the remaining solid dissolved in 100% ethanol to give a final concentration of 10 mM.

The synthetic steroid (50 μM), cofactor (2 mM nicotinamide adenine dinucleotide, NAD) and sodium phosphate buffer containing protease inhibitors to 1 ml total volume were pre-incubated for 15 min at 37 °C in a shaking water bath. One hundred and fifty microlitres of placental protein was subsequently added for 6 h. The reaction was stopped by the addition of 2 ml ice-cold ethyl acetate. Samples were thoroughly mixed and organic and aqueous layers separated. The organic layer containing steroids was dried under high-speed vacuum and steroids were reconstituted in 100 μl of ethyl acetate. Tubes were rotated for at least 30 min, followed by centrifugation at 1000 g for 15 min to increase recovery.

A positive control consisted of [3H]-cortisol (approximately 200 000 cpm per well, Amersham Biosciences, Buckinghamshire, England), 5 μM cortisol and 2 mM NAD in sodium phosphate buffer with protease inhibitors, incubated for 6 h at 37 °C. The negative control consisted of one set of triplicate wells which was incubated for 6 h without protein added, but containing all other components of the reaction (substrate and cofactor).

2.4. Thin-layer chromatography (TLC)

The 100 μl steroid extract was spotted onto aluminium backed TLC plates (Silica Gel 60 F-254, 20 cm x 20 cm, 0.2 μM thickness, Alltech, Deerfield, IL, USA). A marker lane with the steroid and its oxidised metabolite standards were included on each TLC plate. The plates were chromatographed in a glass chamber at room temperature, using mobile phases composed of 95.5 (v/v) or 93.7 (v/v) chloroform:methanol. The developing distance was 8 cm from the starting line.

After mobile phase evaporation the components of interest were visualised using UV light (254 nm). Under these conditions, steroids appeared as black spots against a fluorescent green background. The TLC plates were photographed using a tripod-mounted Minolta 800ui camera with a shutter speed of 1 s and aperture of 5.6 with 400 ASA colour film. Developed photographs were scanned and converted to grayscale for densitometric analysis. Densitometry was used to quantify each band using Scion Image software (version 4.02 for windows, Scion Corporation, Frederick, MD, USA).

2.5. Steroid standards preparation

The oxidised metabolites were prepared according to the method of Shaw and Quinney [34]. Briefly, 20 μg of steroid was incubated for 1 h at room temperature with 1 ml of 50% acetic acid/1% chromium trioxide (w/v). Two hundred and fifty microlitres of dichloromethane was added twice to extract the steroids. The extract was dried over several days under a fume hood and reconstituted in 20 μl of 100% ethanol, rotated for 1 h and centrifuged at 1000 g for 15 min to improve recovery. Preliminary experiments using 1H-cortisol indicated that we could obtain >95% yield of oxidised steroid using this method.

2.6. Statistical analysis

Results are presented as median (interquartile range) or mean ± standard error of the mean, depending on data distribution. Statistical analysis was performed using GraphPad Instat version 3.05 (GraphPad Software, San Diego, CA). ANOVA or the Kruskal–Wallis test was used, along with the appropriate post-hoc tests (Tukey–Kramer multiple comparisons test or Dunn’s multiple comparisons test). P < 0.05 was considered significant.

3. Results

3.1. ICS use for asthma during pregnancy

Table 1 gives the median dose of ICS used in each trimester of pregnancy in the four groups (oral steroids + ICS, beclomethasone alone, budesonide alone and fluticasone alone). ICS dose was significantly lower in the beclomethasone group compared to the oral steroid + ICS group in all trimesters (Kruskal–Wallis test and Dunn’s multiple comparisons test, P < 0.01) and compared to the fluticasone group in the first trimester (Kruskal–Wallis test and Dunn’s multiple comparisons test, P < 0.05). In the first and second trimesters, ICS use was lower in the budesonide group compared to the oral steroid + ICS group (Kruskal–Wallis test and Dunn’s multiple comparisons test, P < 0.01).

3.2. Effect of ICS use on fetal growth and HPA function

Birth weight and placental weight were not significantly different between the groups (ANOVA, P > 0.05). Birth length, head circumference, gestational age at delivery and the birth weight:placental weight ratio were not significantly different between the groups (Kruskal–Wallis test, P > 0.05). Within
Table 1
Dose of ICS used by pregnant women with asthma, and fetal growth and HPA function for each group

<table>
<thead>
<tr>
<th></th>
<th>Non-asthmatic (n = 84)</th>
<th>Oral steroid + ICS (n = 17)</th>
<th>Beclomethasone alone (n = 29)</th>
<th>Budesonide alone (n = 42)</th>
<th>Fluticasone alone (n = 31)</th>
</tr>
</thead>
<tbody>
<tr>
<td>First trimester ICS dose (mg/day)</td>
<td>1000 (800–1600)</td>
<td>100 (0–500) (^{a,b})</td>
<td>300 (0–800)</td>
<td>800 (325–1000)</td>
<td></td>
</tr>
<tr>
<td>Second trimester ICS dose (mg/day)</td>
<td>1000 (800–2000)</td>
<td>400 (100–1000) (^{a})</td>
<td>500 (0–1600) (^{c})</td>
<td>1000 (500–1000)</td>
<td></td>
</tr>
<tr>
<td>Third trimester ICS dose (mg/day)</td>
<td>1000 (1000–2000)</td>
<td>500 (200–1000) (^{d})</td>
<td>800 (650–1600) (^{f})</td>
<td>1000 (500–1000)</td>
<td></td>
</tr>
<tr>
<td>Birth weight (g)</td>
<td>3587 ± 59</td>
<td>3311 ± 150</td>
<td>3582 ± 99</td>
<td>3496 ± 78</td>
<td>3482 ± 120</td>
</tr>
<tr>
<td>Birth length (cm)</td>
<td>52 (51–53) (^{n} (n = 77))</td>
<td>51 (48–52) (^{n} (n = 17))</td>
<td>52 (50–54) (^{n} (n = 28))</td>
<td>52 (50–54) (^{n} (n = 41))</td>
<td>52 (49–54) (^{n} (n = 26))</td>
</tr>
<tr>
<td>Birth head circumference (cm)</td>
<td>35 (34–35.5)</td>
<td>34.5 (33.5–35)</td>
<td>34.5 (34–36)</td>
<td>35 (33–36)</td>
<td>34.5 (33–36)</td>
</tr>
<tr>
<td>Placental weight (g)</td>
<td>628 ± 25 (^{n} (n = 40))</td>
<td>622 ± 46 (^{n} (n = 6))</td>
<td>654 ± 53 (^{n} (n = 10))</td>
<td>710 ± 46 (^{n} (n = 12))</td>
<td>621 ± 35 (^{n} (n = 20))</td>
</tr>
<tr>
<td>Birth weight/placental weight ratio</td>
<td>5.7 (5.3–6.2) (^{n} (n = 40))</td>
<td>5.6 (5.4–6.5) (^{n} (n = 6))</td>
<td>6.1 (4.9–6.9) (^{n} (n = 10))</td>
<td>5.3 (4.8–6.1) (^{n} (n = 12))</td>
<td>5.5 (5.1–5.9) (^{n} (n = 20))</td>
</tr>
<tr>
<td>Gestational age at birth (weeks)</td>
<td>40 (39–40.8)</td>
<td>39 (38–41)</td>
<td>39.3 (38–40)</td>
<td>39.9 (39–40.8)</td>
<td>39.9 (39–41)</td>
</tr>
<tr>
<td>% Vaginal delivery</td>
<td>85</td>
<td>100</td>
<td>79</td>
<td>71</td>
<td>68</td>
</tr>
<tr>
<td>% Caeasarean section</td>
<td>15</td>
<td>0</td>
<td>21</td>
<td>29</td>
<td>32</td>
</tr>
<tr>
<td>Umbilical vein cortisol (nmol/l)</td>
<td>195 ± 14 (^{n} (n = 24))</td>
<td>198 ± 25 (^{n} (n = 10))</td>
<td>324 ± 49 (^{d} (n = 19))</td>
<td>191 ± 40 (^{n} (n = 10))</td>
<td>246 ± 35 (^{n} (n = 13))</td>
</tr>
<tr>
<td>Umbilical vein estriol (nmol/l)</td>
<td>731 (545–903) (^{n} (n = 16))</td>
<td>572 (496–886) (^{n} (n = 7))</td>
<td>791 (746–1020) (^{n} (n = 17))</td>
<td>976 (708–1048) (^{n} (n = 6))</td>
<td>766 (704–841) (^{n} (n = 7))</td>
</tr>
</tbody>
</table>

Values are median (interquartile range) or mean ± standard error of mean or percentage of subjects in each group.

ICS = inhaled corticosteroid.

- Indicates P < 0.01 beclomethasone alone vs oral steroid + ICS (Kruskal–Wallis test and Dunn’s multiple comparisons test).
- Indicates P < 0.05 beclomethasone alone vs fluticasone alone (Kruskal–Wallis test and Dunn’s multiple comparisons test).
- Indicates P < 0.01 budesonide vs oral steroid + ICS (Kruskal–Wallis test and Dunn’s multiple comparisons test).
- Indicates P < 0.05 beclomethasone alone vs non-asthmatic (ANOVA and Tukey–Kramer multiple comparisons test).

Each group, there were no significant differences between male and female neonates (data not shown, P > 0.05). There was one preterm delivery in the control group, one in the budesonide group, one in the fluticasone group and one in the oral steroid + ICS group.

Fetal HPA function was assessed by measuring umbilical vein cortisol and estriol in a subset of samples. This subset was similar to the whole group in terms of ICS use and fetal growth parameters (data not shown, P > 0.05). There were no differences in umbilical vein estriol concentration between the groups (Kruskal–Wallis ANOVA, P > 0.05), while umbilical vein cortisol was significantly higher in the beclomethasone group compared to the non-asthmatic group (ANOVA, P = 0.023, Tukey–Kramer multiple comparisons test, P < 0.05).

3.3. Metabolism of synthetic steroids by the placenta

The metabolism of the synthetic steroids was monitored over time using the TLC method (Fig. 2). The formation of the 11-keto metabolite of beclomethasone was first observed at 60 min (Fig. 2a). Other placental metabolites were also present. No 11-keto metabolite was observed for budesonide (Fig. 2b) or fluticasone (Fig. 2c). The 11-keto metabolite of prednisolone was first observed at 240 min (Fig. 2d), the 11-keto metabolite of dexamethasone from 120 min (Fig. 2e) and the 11-keto metabolite of betamethasone from 60 min (Fig. 2f). Separation of betamethasone, dexamethasone and prednisolone from their metabolites was not sufficient under the conditions chosen to investigate the presence of additional metabolites.

Densitometric analysis indicated that at 6 h, 11.6% ± 1.0% of beclomethasone was metabolised by placental 11β-HSD2 into the 11-keto derivative (Fig. 3), while an additional 21.9% ± 0.8% was metabolised into unknown steroid metabolites, represented by three additional bands on the TLC plate (Fig. 2a). No budesonide or fluticasone metabolites were detected using the TLC/densitometry method. 23.4% ± 1.3% of prednisolone was metabolised by 11β-HSD2 into the 11-keto derivative and 13.0% ± 1.4% of dexamethasone and 11.6% ± 0.5% of betamethasone were metabolised into 11-keto derivatives (Fig. 3). Metabolism of prednisolone and betamethasone by 11β-HSD2 was significantly greater than beclomethasone dipropionate and dexamethasone (Fig. 3, ANOVA and Tukey–Kramer multiple comparisons test, P < 0.001).

4. Discussion

In this study, we have demonstrated using an in vitro enzyme assay and TLC method that placental 11β-HSD2 metabolises the synthetic steroids beclomethasone, prednisolone, dexamethasone and betamethasone, but not budesonide or fluticasone. This is the first study to examine placental metabolism of beclomethasone, budesonide and fluticasone, drugs commonly used to treat pregnant women with asthma. We have previously measured placental 11β-HSD2 activity (using cortisol as the substrate) in samples collected from pregnant women with and without asthma [22]. When these data were reanalysed based on oral or inhaled steroid use, there were no significant differences between the groups (data not shown).

Previous studies of placental metabolism of synthetic steroids have been conflicting, possibly due to differences in techniques used to assess enzyme activity. The four main techniques which have been used are incubation of placental minces with steroid [35,36], incubation of placental microsomal protein with steroid [37], perfusion of the placenta with steroids with subsequent measurement of metabolites in the perfusate [36,38] and in vivo administration of steroids to
pregnant women with measurement of metabolites in cord plasma or amniotic fluid [39,40]. Using an in vitro method where minces of placental tissue were incubated for 2 h with steroid, Blanford and Murphy found that 67% of cortisol and 51% of prednisolone were metabolised, while only 2% of dexamethasone and 7% of betamethasone were converted to their 11-keto metabolites [35]. Another study found that the conversion of dexamethasone and betamethasone to their 11-keto metabolites was higher when using a perfused placenta method as opposed to one with minced placental pieces [36]. In this study, the metabolisms of cortisol, prednisolone, dexamethasone and betamethasone were similar [36]. Some studies have used in vivo techniques. For example, Anderson et al. measured betamethasone in maternal plasma, umbilical cord plasma and amniotic fluid, following 3 consecutive days of betamethasone administration to the mother [40]. They found that betamethasone was transferred to the fetus and was measurable in amniotic fluid and umbilical cord blood at a similar concentration to that found in maternal plasma [40], indicating that there was very little placental metabolism of betamethasone. Our in vitro technique was similar to that used in other recent studies and involved purifying 11β-HSD2 protein from placental microsomes and incubating this preparation with the steroids of interest. This technique gives greater enzyme activity than more crude homogenates such as placental minces. Using our in vitro technique, 25% of prednisolone was converted to prednisone during a 6-h incubation of placental microsomes. An in vivo study of the transplacental passage of prednisolone, where concentrations were measured in maternal and fetal plasma following intravenous infusion, found that concentrations of prednisolone in the fetus were 8–10-fold lower than the mother and the 11-keto metabolite (prednisone) was at a similar concentration in both mother and fetus [39]. This suggested significant metabolism of prednisolone by the placental 11β-HSD2 barrier. The results of our study show that 11β-HSD2 metabolism of prednisolone and betamethasone is significantly higher than that of beclomethasone and dexamethasone, while there was no measurable metabolism of budesonide or fluticasone by the placenta.

There are limitations to the technique we used to measure metabolism. Because of the need to quantify the steroids and
their metabolites on the TLC plate, a large concentration of steroid substrate was used (50 μM), which was 10-fold higher than the saturating concentration we previously used in a similar study of placental 11β-HSD2 activity [22]. The use of a more efficient separation technique such as high performance liquid chromatography (HPLC) would improve the results obtained from our study. However, other techniques are limited by the detection systems and the availability of the oxidised steroid compounds. In addition, it would be helpful to measure the steroids and their metabolites in the umbilical cord blood to determine whether they are likely to reach the fetal circulation. It is not known whether physiologically active concentrations of the steroids used to treat asthma reach the fetus. However, it is also known that use of these medications in adults can result in systemic side effects such as adrenal suppression [41] and reduced bone mineral density in women [42]. It is possible that in women who use high doses of inhaled or oral steroids during pregnancy, there may be some transfer of these compounds to the fetal circulation. Metabolism of these steroids by placental 11β-HSD2 may be a mechanism by which the fetus is protected from their adverse effects.

We found that there was more metabolism of prednisolone and betamethasone compared to beclomethasone and dexamethasone, while no metabolism of budesonide and fluticasone was detected. This could be due to the different chemical structures of these steroids. Budesonide and fluticasone both contain bulky side groups attached to carbons 15 and 16 (budesonide) and carbon 17 (fluticasone). In addition, they are the only steroids of this group without a hydroxyl group on carbon 17. These features may inhibit their binding to 11β-HSD2. Betamethasone and dexamethasone are stereoisomers, with only the methyl group on carbon 16 in different positions. It is possible that the position of this methyl group leads to altered binding to the enzyme. There are also differences in the halogen molecule on carbon 9. Beclomethasone has a chlorine attached, while betamethasone, dexamethasone and fluticasone have a fluorine atom. The natural substrate, cortisol, as well as prednisolone do not contain a halogen in this position. It has been previously suggested that the 9-halogen group provides protection from metabolism by 11β-HSD2 [43,44].

In this study, we have found no significant effect of oral or inhaled steroid use for asthma treatment during pregnancy on fetal growth parameters and gestational age at birth, in a relatively small group of women. In addition, there was no effect of ICS use on fetal estriol concentrations, suggesting that these steroids do not alter fetal HPA function. We did, however, find a significant increase in fetal cortisol concentration in the group using beclomethasone, although this was a modest increase compared to the non-asthmatic control group only.

There is extensive evidence for the safety of the major drug classes used to treat asthma during pregnancy, including ICS [7,45,46]. A recent Swedish study found that the use of budesonide during pregnancy does not affect gestational age, birth weight, birth length or the rate of still-births or multiple births [7]. This data came from 2968 women who used inhaled budesonide during pregnancy and was compared to 7719 women who used asthma medications other than steroids, and a control population of over 293,000 women. The safety of oral steroids for asthma during pregnancy is less clear, with two large prospective cohort studies recently finding an association between oral steroid use and preterm delivery [2,4]. Information is lacking on the safety of some of the newer ICS drugs such as fluticasone propionate. In this study we found no difference in size at birth between asthmatic women using beclomethasone, budesonide or fluticasone alone and ICS taken with periodic oral steroids. Recently, Namazy et al. studied asthmatic women using beclomethasone, budesonide, fluticasone, triamcinolone or flunisolide and there was no increase in the rate of small for gestational age infants compared to population averages [8].

We conclude firstly, that the use of the inhaled steroids, beclomethasone, budesonide and fluticasone, for asthma treatment during pregnancy does not result in any significant effects on neonatal size at birth. Secondly, in women without asthma, the placenta metabolises the synthetic steroids beclomethasone, prednisolone, dexamethasone and betamethasone, but does not metabolise budesonide or fluticasone.

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